

アプリケーションノート

## Quantification of Morphine, Morphine-3-glucuronide and Morphine-6-glucuronide in Biological Samples by LC-MS/MS

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*For forensic toxicology use only.*

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## Abstract

In this application note, we present a sensitive method for the quantification of morphine and its glucuronide metabolites. The method involves a simple SPE purification prior to analysis using LC/MRM and is suitable for plasma, whole blood or urine samples.

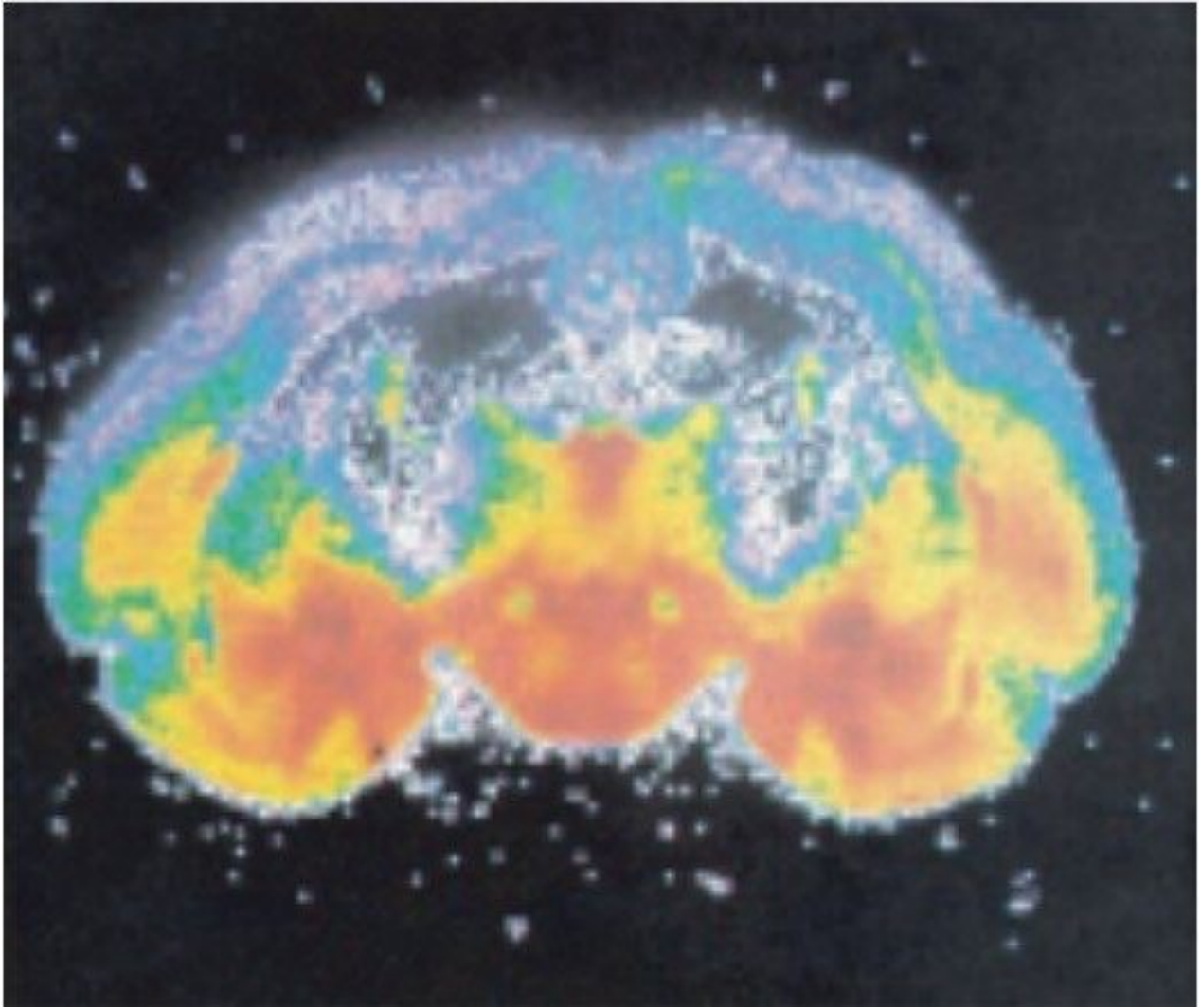
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## Introduction

Morphine is a potent analgesic isolated from the opium poppy *papaver somniferum* and traditionally used for the treatment of moderate to severe pain. Analgesia results from the action of morphine at the opioid receptors of the spinal cord and brain (Figure 1), where it attenuates both the speed of the impulse and the perception of pain.

In human subjects, morphine is extensively metabolised (primarily by conjugation with glucuronic acid) to form morphine-3-glucuronide (M3G) and morphine-6-glucuronide (M6G). Whilst, the principal metabolite i.e. M3G, has little or no analgesic effect, M6G has been shown to be highly effective and is believed likely to contribute significantly to the overall effectiveness of morphine.<sup>1</sup> Hence, quantification of both the parent drug and metabolites is desirable for pharmacokinetic studies.

Previously we have described a LC-MS/MS method that allows the quantification of morphine and several other opiates in urine.<sup>2</sup> Here we present a simple method that enables the quantification of morphine in plasma, whole blood and urine. Furthermore this procedure allows differentiation between two isobaric glucuronide metabolites.



*Figure 1. Image of guinea-pig brain. The red areas represent the highest density of opioid receptors; yellow areas represent moderate density; whilst blue, purple and white represent low density.*

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## Experimental

### Sample Preparation

Biological samples were prepared for LC-MS/MS analysis by means of a simple, solid-phase extraction (SPE) procedure. A Waters Oasis HLB extraction Cartridge (1 cc/30 mg) was firstly conditioned with 1 mL volumes of each of the following: methanol, water and ammonium carbonate (10 mM, pH 8.8). Samples (100  $\mu$ L, spiked with deuterated internal standards) were made up to a final volume of 1 mL with ammonium carbonate before applying to the pre-conditioned cartridge. The cartridge was then washed with 1 mL

ammonium carbonate before elution of the sample using 100% methanol (0.5 mL). Eluents were dried using a Savant Speedvac Plus evaporator and then redissolved in 100  $\mu$ L of mobile phase. Reconstituted samples were briefly vortex mixed before the analysis of 10  $\mu$ L using LC in conjunction with multiple reaction monitoring (MRM).

## LC-MS/MS

A Waters Quattro micro triple quadrupole mass spectrometer fitted with ZSpray ion interface was used for all analyses. Ionisation was achieved using electrospray in the positive ionisation mode (ES+). Details of the MRM conditions are given in Table 1.

Compound	Precursor ion (m/z)	Product ion (m/z)	Cone Voltage (V)	Collision energy (eV)
Morphine	286	165	45	38
Morphine-d3	289	165	45	40
Morphine-M3G-glucuronide	462	286	45	28
Morphine-M3G-d3-glucuronide	465	289	45	30
Morphine-M6G-glucuronide	462	286	45	28

Table 1. MRM transitions and conditions for the measurement of morphine and its metabolites. The deuterated analogues of morphine and morphine-3-glucuronide were also included for the purpose of internal standardisation.

LC analyses were performed using a Waters 2795 separations module. Chromatography was achieved using a C<sub>18</sub> column (3.9 x 150 mm) eluted isocratically with 0.1% formic acid:acetonitrile (97:3) at a flow rate of 0.3 mL/min. Column temperature was maintained at 30 °C. All aspects of system operation and data acquisition were controlled using MassLynx 4.0 software with automated data processing using the QuanLynx program.

## Results and Discussion

A series of calibrators (0.5-500  $\mu$ g/L) were prepared in duplicate by adding standards to blank plasma, whole blood or urine. Samples were then extracted using the SPE method described above prior to LC/MRM analysis.

Following LC/MRM analysis, the areas under the specific MRM chromatograms were integrated.

Figure 2 shows the extracted MRM chromatogram of morphine, M3G and M6G obtained with a 10  $\mu$ L injection of the 5  $\mu$ g/L plasma calibrator. Opiates were quantified by reference to the integrated area of the deuterated internal standards. Responses were linear ( $r = >0.999$ ) over the range investigated for all 3 compounds and in each matrix (Figure 3 shows a typical standard curve for M3G in urine).

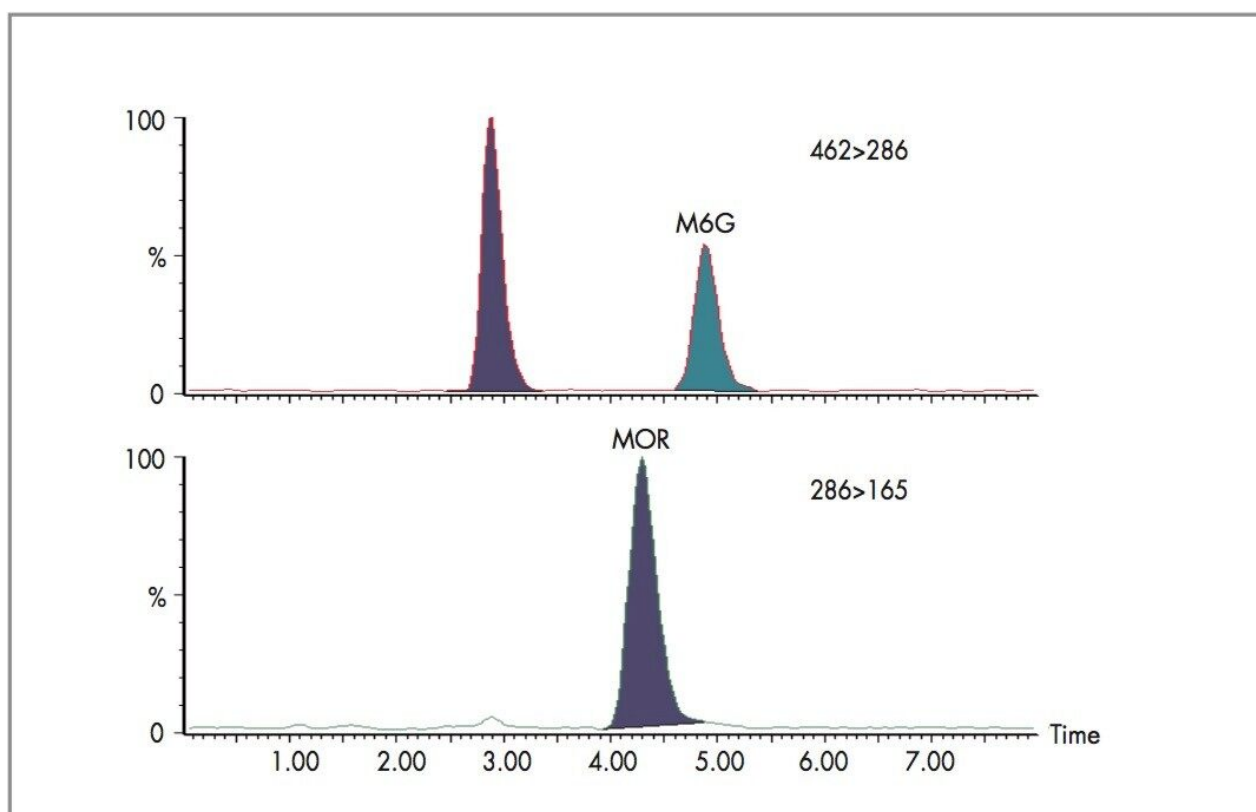


Figure 2. MRM chromatogram for morphine (MOR), M3G and M6G. The above responses were obtained with a 10  $\mu$ L injection of the 5  $\mu$ g/L plasma calibrator. Due to the isobaric nature of M3G and M6G chromatographic resolution is required to enable identification.

Compound name: Morphine-3-glucuronide  
Correlation coefficient:  $r = 0.999883$ ,  $r^2 = 0.999766$   
Calibration curve:  $0.09404 * x + 0.100968$   
Response type: Internal Std ( Ref 4 ), Area \* ( IS Conc. / IS Area )  
Curve type: Linear, Origin: Exclude, Weighting: 1/x Axis trans: None

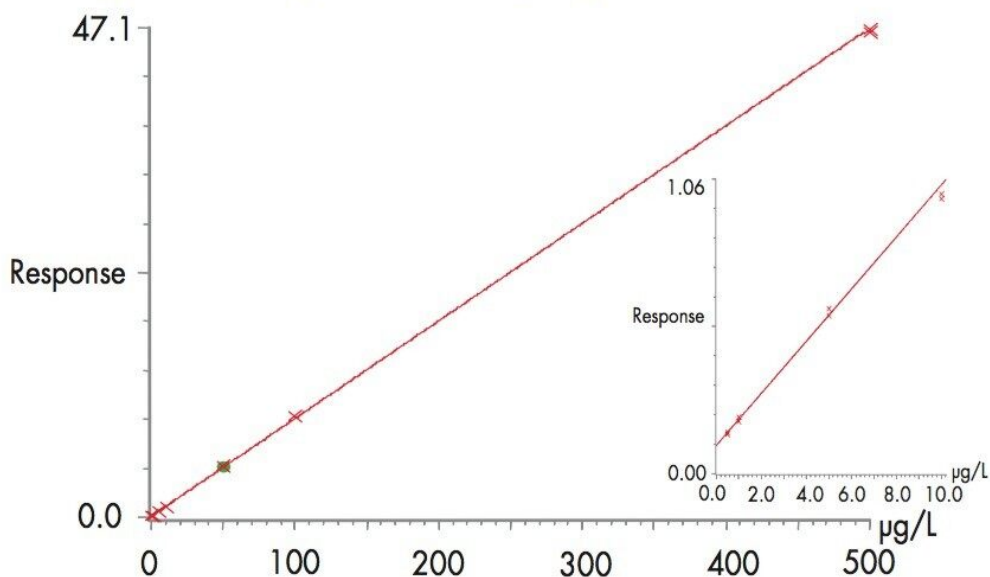


Figure 3. Standard curve for M3G in urine. Responses (duplicates) were calculated in reference to the integrated area of the deuterated internal standards. The inserted figure shows the response for the range 0-10 µg/L.

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## Conclusion

We present a sensitive method for the quantification of morphine and its glucuronide metabolites. The method involves a simple SPE purification prior to analysis using LC/MRM and is suitable for plasma, whole blood or urine samples.

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## References

1. The Analgesic Effect of Morphine-6-Glucuronide. R Osborne, P Thomson, S Joel, D Trew, N Patel and M Slevin. *Br J Clin. Pharmacol.* 1992. 34 (2) 130-8.

2. Opiates: Use or Abuse? Quantification of Opiates in Human Urine. (Waters Application Brief WAB45). M Wood, K Rush\*, M Morris and A Traynor\*. Clinical Applications Development Group, UK Limited, Manchester, UK.

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