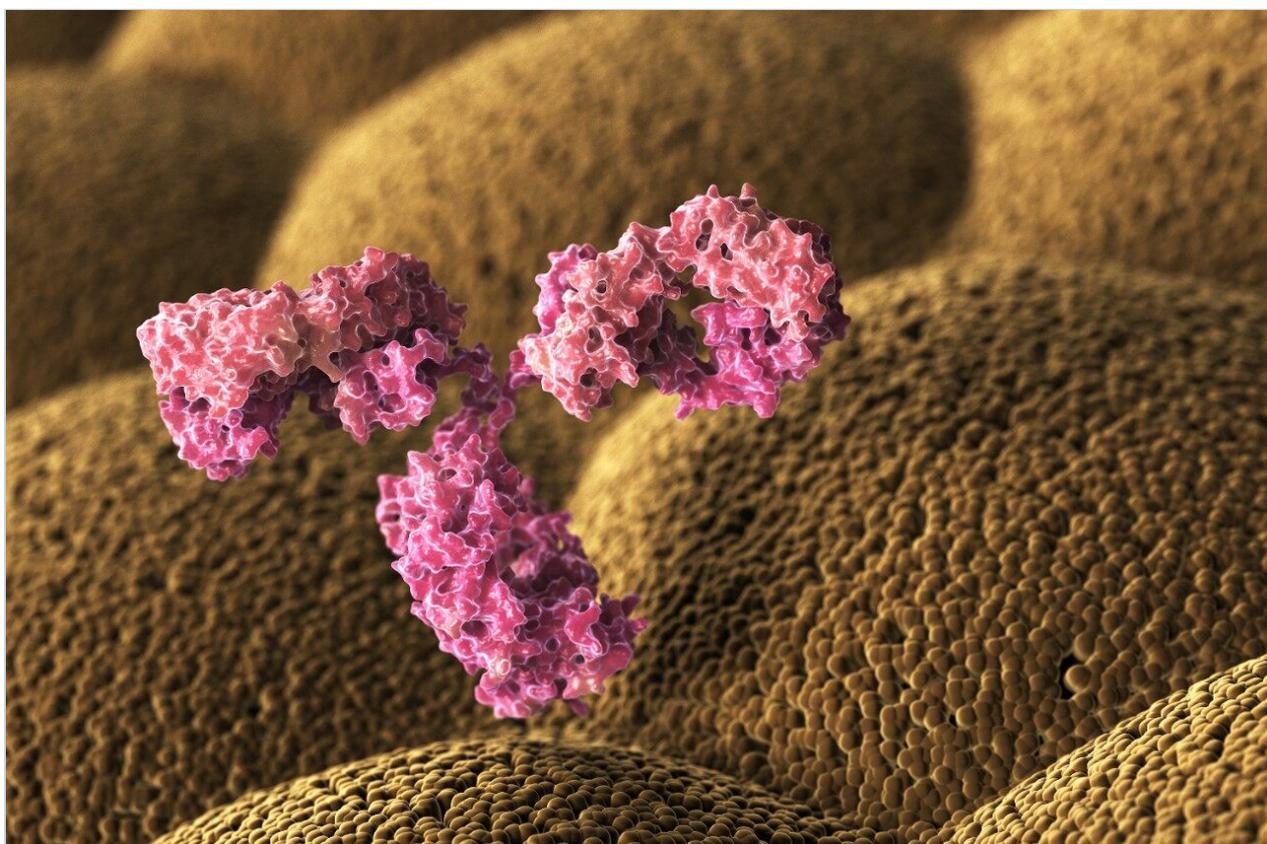


Generic Protein and Peptide Level Sample Preparation for Protein Bioanalysis

日本ウォーターズ株式会社



This is an Application Brief and does not contain a detailed Experimental section.

Abstract

This application brief demonstrates to evaluate the impact of various degrees of sample preparation on the

quantification of therapeutic or diagnostic antibodies in plasma.

Benefits

Answers these questions: What is the industry pressure that requires this application? Why is this specific analytical capability needed?

Introduction

Increasingly, new drugs in development are based on large biomolecules such as peptides and proteins as opposed to traditional small molecule entities. This shift to therapeutic “biopharmaceuticals” has led to an increased need for appropriate bioanalytical strategies. Traditional ligand binding assays (LBA) such as ELISA require individual reagents that can take up to six months to develop, often at significant expense. By contrast, LC-MS/MS assays have the advantage of short development times, the ability to multiplex different analytes, and are based upon a well established technology with which many investigators are comfortable. A significant challenge, however, is the complexity of the plasma proteome and the fact that many biotherapeutics are present at concentrations many orders of magnitude lower than the most abundant, endogenous plasma proteins. These analytical challenges make the need for appropriate sample preparation paramount. Figure 1 shows the typical steps in common protein bioanalysis workflows, including cleanup at the protein level using techniques such as affinity purification or depletion plates, and post-digest peptide level cleanup options such as SPE.

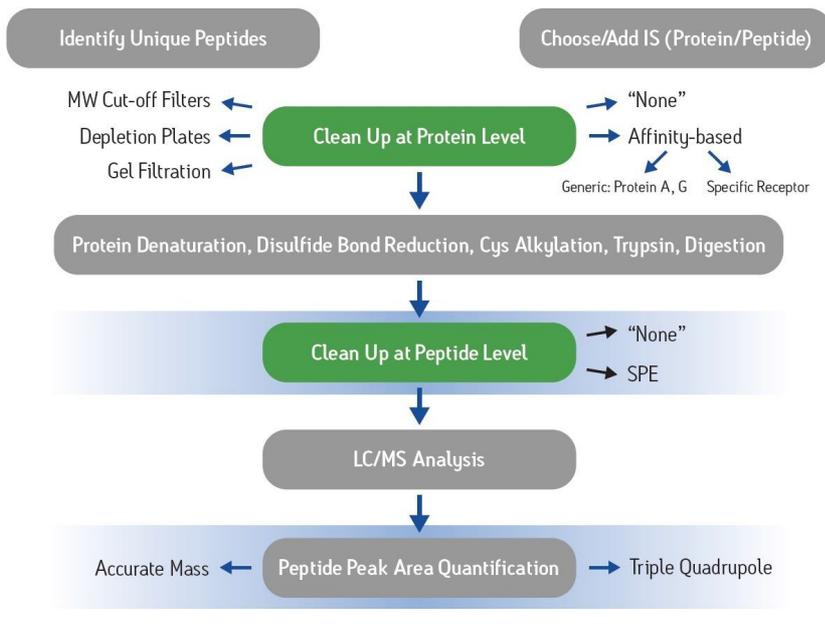


Figure 1. Common steps in the protein bioanalysis workflow. For this specific example, protein A cleanup was followed by trypsin digestion and mixed mode SPE cleanup.

Results and Discussion

To demonstrate the advantages of appropriate sample preparation, the therapeutic monoclonal antibody trastuzumab was analyzed in human plasma. Two parallel strategies were used. In one scheme, 96-well protein A purification plates were used to isolate the IgG fraction (including trastuzumab) from plasma samples. In a second scheme, no protein level cleanup was performed. Each set of samples was then denatured with *RapiGest*, reduced, alkylated and digested with trypsin to yield peptide fragments suitable for LC-MS/MS analysis. These resulting digests were then split into two aliquots. One aliquot was directly analyzed by LC-MS/MS while a second was further cleaned up using Oasis MCX μ Elution plates prior to LC-MS/MS analysis.

Figure 2 demonstrates the differences and incremental benefits achieved through various degrees of sample preparation. The sample in Figure 2A was a direct digestion of the plasma, followed by direct injection of the subsequent digest. There was no sample cleanup performed at either the protein or peptide level. The resultant chromatogram yields a poor intensity peak which co-elutes with closely related interferences, rendering accurate quantification challenging and yielding a poor LLOQ. The sample in Figure 2B was cleaned up at the protein level only using protein A 96-well plates. This type of sample prep is

generic enough to be applied in any workflow where the drug is an IgG-based antibody, making it ideal for a discovery setting. Furthermore, the 96-well format is also compatible with high-throughput analysis. The increase in peak area for the signature peptide relative to no protein-level isolation (Figure 2A) is nearly 5 fold. The chromatogram in Figure 2C represents a sample which has been cleaned up both at the protein level with protein A and at the peptide level with mixed-mode cation exchange SPE. The incremental benefit over protein A cleanup alone (Figure 2B) is additional 2 fold increase in peak area and an elimination of adjacent peaks, increasing specificity of the assay. Mixed-mode SPE cleanup not only removes interfering peptides, but also removes digest reagents, buffers, and other plasma components such as phospholipids, improving instrument robustness and data quality. Protein A sample preparation also eliminates high abundance plasma proteins such as albumin and transferrin, which reduces the required SPE capacity and facilitates the use of the μ Elution plate format. The major benefit of this SPE format is concentration of the sample without evaporation, thus minimizing peptide losses due to adsorption. Although not tested here, mixed-mode SPE can also be used to purify signature peptides from direct plasma digests (without protein-level cleanup). However, a larger bed mass 96-well plate may be required to accommodate the same sample volume due to the increased concentration of undesired peptides. An additional advantage to the use of generic protein level isolation, such as protein A, is that it reduces the required amount of trypsin by almost 10X, significantly reducing the assay cost.

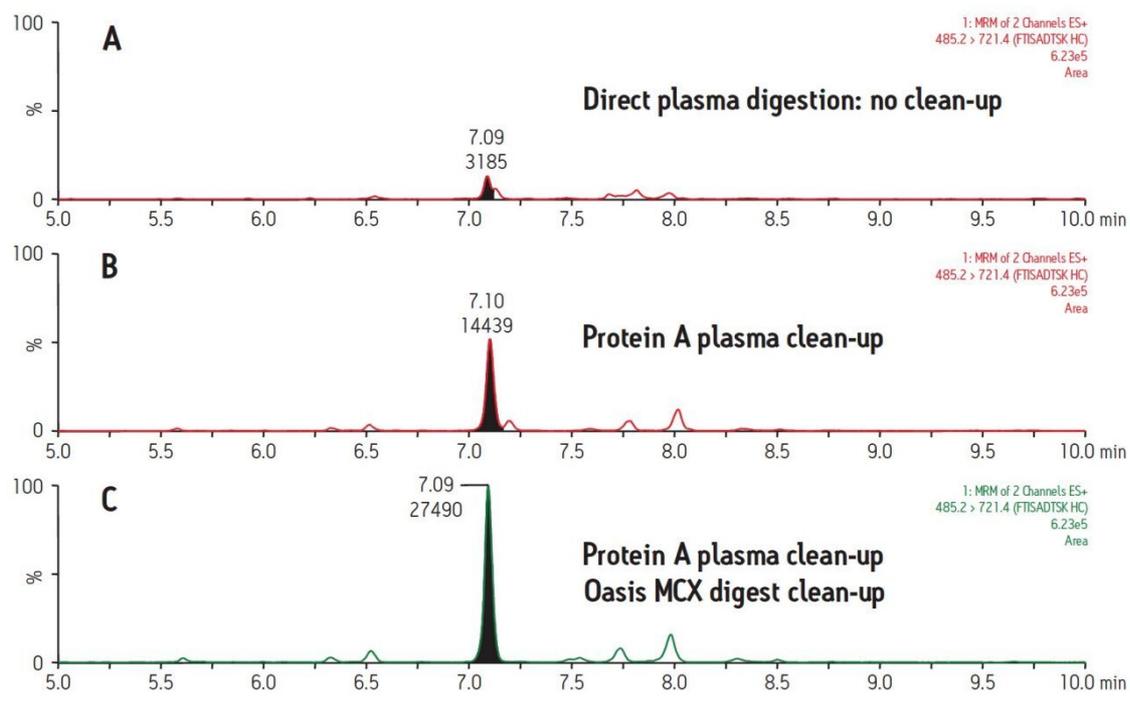


Figure 2. Comparison of sample preparation options for protein quantification.

Conclusion

- Generic protein-level cleanup, such as protein A, significantly reduces sample complexity by eliminating nearly 85% of endogenous proteins, improving detection limits by 5X in this assay. Protein A fractionation also minimizes assay cost by reducing the amount of trypsin required.
- Oasis mixed mode MCX μ Elution plates can be used to further improve sensitivity, specificity, and assay and system robustness for the bioanalysis of therapeutic proteins.
 - The mixed-mode sorbent and μ Elution format enables the selective cleanup and concentration of specific signature peptides without evaporation, minimizing potential losses of desired peptides due to adsorption.
 - SPE of protein digests also eliminates digest reagents, buffers, and other plasma components such as phospholipids.
 - In conjunction with other Oasis sorbents (WCX, MAX, WAX, and HLB), the technique is generic enough to enable cleanup of a wide variety of therapeutic or diagnostic proteins after digestion in both the drug discovery and development settings.

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