

Note d'application

## Chiral Purification of Volatile Flavors and Fragrances by SFC

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Abstract

In this application note, linalool and terpinen-4-ol were used to demonstrate the chiral purification capabilities of SFC for volatile flavor and fragrance compounds. First, a recovery study was performed by collecting the enantiomers from ( $\pm$ )-terpinen-4-ol and ( $\pm$ )-linalool standards. These compounds were then enantiomerically purified from lavender and tea tree essential oils. Stacked injections and fast chiral separations reduced the time required for the enantiomeric purification of flavor and fragrance compounds.

## Benefits

- The Investigator SFC System allows for simplified chiral separations and for the purification and recovery of volatile compounds without the need for the complex sample trapping that is necessary in preparative gas chromatography (GC).
- SFC has a much higher loading capacity and shorter run times than GC.
- Supercritical fluid chromatography (SFC) uses lower chromatographic operating temperatures, which are more suitable for volatile and unstable compounds to improve upon the low recoveries inherent in flavor and fragrance purification.
- The SFC chiral purification method uses non-toxic ethanol and CO<sub>2</sub> as the mobile phase, which is important when purifying compounds for human use or possible consumption.
- The Investigator SFC System improves collection efficiency by using stacked injections, resulting in higher throughput.

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## Introduction

Found in foods, wines, spices, perfumes, and essential oils, flavors and fragrances are abundant in nature and enhance life.<sup>1</sup> In general, these compounds are small molecules that are sufficiently volatile to be sensed through taste or smell. They cover a wide range of chemicals, including terpenes, phenols, aldehydes, and esters, many of which are chiral.<sup>2</sup> By obtaining them through natural sources, chemical synthesis, or fermentation, the flavor and fragrance industry ensures a constant infusion of new experiences to surprise and please the chemical senses.<sup>1-2</sup>

The interaction of compounds with biological systems has long been shown to be stereoselective.<sup>3</sup> Just as enantiomers of chiral drugs exhibit different pharmacological activity, chirality also plays an important role in the flavor and fragrance chemistry.<sup>4</sup> These chiral enantiomers dictate not only taste or odor quality, but also intensity. Therefore, chemists devote great effort to investigate enantiopure chiral flavors and fragrances.<sup>5</sup>

Care is taken to make sure only the desired odor or flavor active isomer is added to reduce toxicological risks and, specifically, to meet strict regulations in the food and beverage industries. To that end, highly pure compounds are required, and no toxic chemicals are permitted during preparation.<sup>2</sup>

Many flavor and fragrance compounds are purified by simple fractional distillation. However, this technique does not have the same selectivity as chromatographic methods, nor can it distinguish between chiral enantiomers. Currently, the most common method for purifying flavor and fragrance compounds is by prep GC. While this technique exhibits high resolution for these types of volatile compounds, purification by GC can be quite challenging. GC exhibits low loading capacities due to the limitations of capillary columns, as well as long run times, often resulting in only a few micrograms being collected over several hours. Also, the high temperatures typical of GC increase the risk of sample loss and degradation. To enable fraction collection by GC, cooled (often with liquid nitrogen) traps are required, and multiple traps are needed if there is more than one compound of interest in a sample.<sup>6</sup>

SFC is a chromatographic technique that employs compressed CO<sub>2</sub> as the main component of the mobile phase. In contrast to GC, packed columns and low temperatures improve column loading and compound stability. A simple multiport valve (for easy collection of multiple peaks) and a make-up pump simplify fraction collection without the need for special trapping equipment.

In this application note, linalool and terpinen-4-ol (Figure 1) were used to demonstrate the chiral purification capabilities of SFC for volatile flavor and fragrance compounds. First, a recovery study was performed by collecting the enantiomers from (±)-terpinen-4-ol and (±)-linalool standards. These compounds were then enantiomerically purified from lavender and tea tree essential oils. Stacked injections and fast chiral separations reduced the time required for the enantiomeric purification of flavor and fragrance compounds.

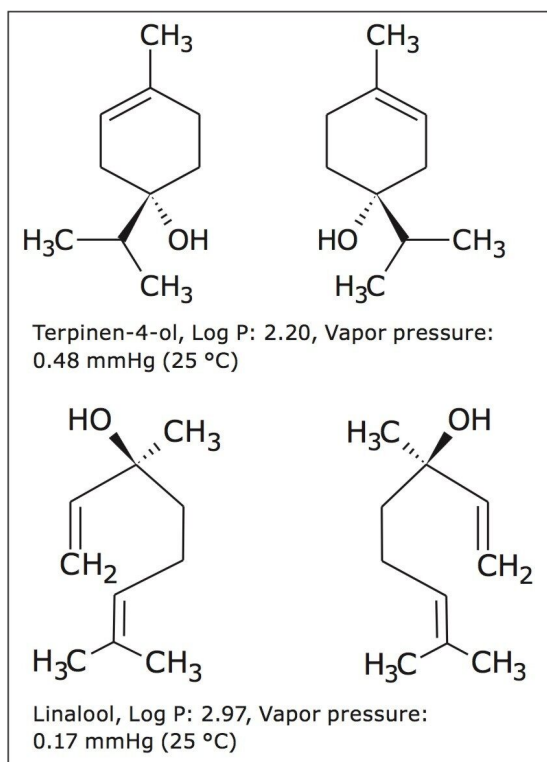


Figure 1. Chemical structures, Log P, and vapor pressure values for terpinen-4-ol and linalool.

## Experimental

### Sample description

All recovery and method development samples were made up at concentrations of approximately 10 mg/mL in ethanol. Collections were performed in sets of ten 100- $\mu$ L stacked injections, for a total of 1 mL injected. The recoveries were performed in triplicate and all collected enantiomeric fractions were transferred to 25-mL volumetric flasks and brought up to volume in ethanol. Re-analysis standards were made by diluting 1 mL (the total injection volume) of the 10-mg/mL solutions in 25 mL ethanol. Due to the fairly small percentage of the desired compound in the tea tree and lavender essential oils, these samples ultimately required higher concentrations of 50 mg/mL and 30 mg/mL, respectively.

### Preparative SFC conditions

SFC system:	Waters Investigator SFC System
Preparative column:	CHIRALPAK AD-H, 5 $\mu$ m, 10 x 250 mm
Mobile phase A:	CO <sub>2</sub>
Mobile phase B:	Ethanol
Make-up solvent:	Ethanol
Flow rate:	12 mL/min
Additional conditions:	Recorded in Table 1

	(±)-Terpinen-4-ol	(±)-Linalool	Tea tree oil	Lavender oil
<b>%B (Isocratic)</b>	10	15	8	18
<b>BPR pressure</b>	200 Bar	120 Bar	120 Bar	120 Bar
<b>Oven temperature</b>	35 °C	35 °C	30 °C	35 °C
<b>Make-up flow</b>	2 mL/min	2 mL/min	2 mL/min	1.5 mL/min
<b>HE temperature</b>	35 °C	25 °C	30 °C	25 °C
<b>Sample concentration</b>	10 mg/mL	10 mg/mL	50 mg/mL	30 mg/mL
<b>Injection volume</b>	100 $\mu$ L	100 $\mu$ L	100 $\mu$ L	100 $\mu$ L

Table 1. Preparative SFC method conditions.

## Fraction analysis

Fraction analysis was performed on the same Investigator SFC system, using a 5  $\mu$ m, 4.6 x 250 mm CHIRALPAK AD-H Column. For the recovery study, the areas for the diluted enantiomeric fractions were compared against the areas for the re-analysis standards. The undiluted fractions from the essential oils were tested for %purity against all impurities (enantiomeric or matrix).

## PDA/UV conditions

Detector: 2998 PDA Detector

Scan: 220–300 nm

Collection: Single wavelength 220 nm

## Data management

ChromScope v1.2

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## Results and Discussion

There are many challenges inherent in the chiral purification of volatile flavor and fragrance compounds. The first challenge is in the chromatography itself; not only do the compounds have to be resolved achirally from the matrix, but enantiomeric resolution is also required. For purification, good chromatographic resolution enables higher column loading, improved product purity, and efficiency. Because of the volatile and instable nature of these compounds, analysis and purification must be accomplished without significant sample loss due to evaporation or degradation. Finally, the preparation of these compounds requires the use of non-toxic chemicals, especially if the products are destined for human use or consumption.

SFC uses CO<sub>2</sub> as the primary component of the mobile phase, and in most cases alcohol as solvent B. In this case, ethanol is used due to its non-toxic nature and compatibility with not only flavor and fragrance analytes, but also with any post purification processes down-stream. Also, due to the evaporation of the CO<sub>2</sub> after collection, there is much less solvent collected than in traditional LC techniques. Another advantage of SFC for the purification of flavor and fragrance compounds is the relatively mild conditions used for separation compared to GC or HPLC. Low temperatures and lack of additives decrease the possibility of evaporation and degradation of the target compounds during analysis and purification.

### Recovery study

Enantiomeric separations of the linalool and terpinen-4-ol isomers were achieved under isocratic conditions, on the 5 µm, 10 x 250 mm CHIRALPAK AD-H Column in less than 4 minutes. High resolution and good peak shape allowed for loading of 1 mg per injection. Based on this chromatography, much more could have been loaded, but for the purposes of the recovery study, this loading was ideal. Isocratic method conditions made it possible to do stacked injections, resulting in the injection and collection of approximately 10 mg in less than 30 minutes (figure 2) for both terpinen-4-ol and linalool chiral purifications.

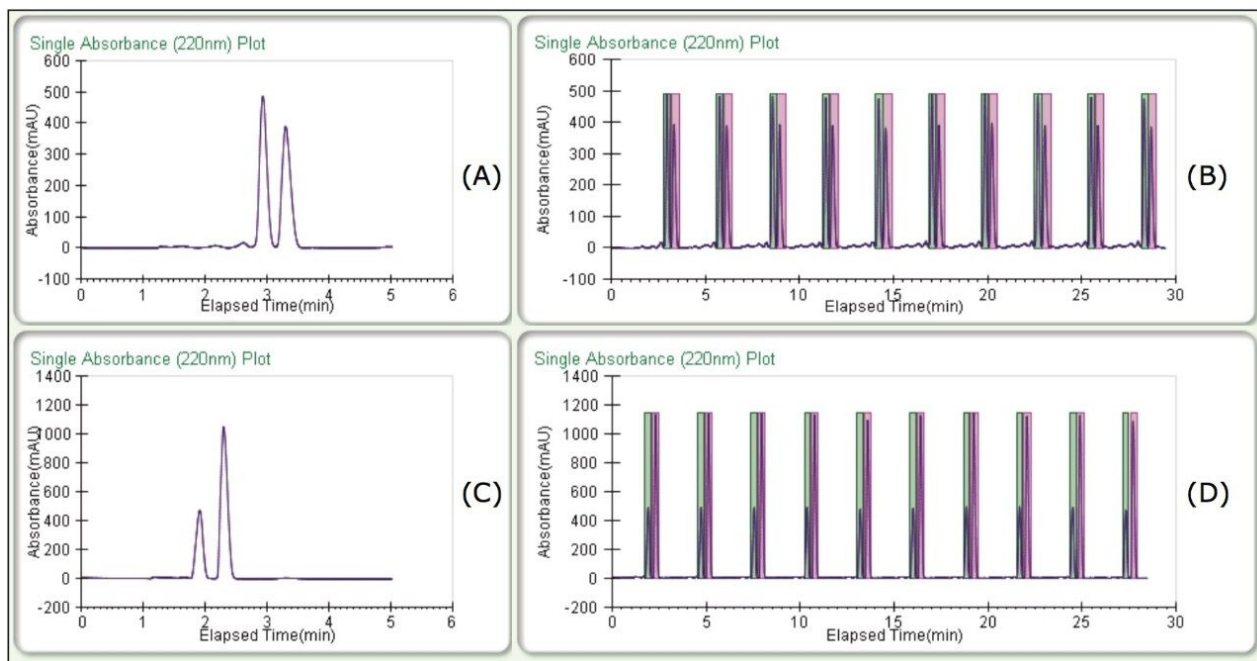


Figure 2. Preparative separations and stacked collections for (±)-terpinen-4-ol (A and B) and (±)-linalool (C and D).

As part of the recovery study, different heat exchanger (HE) temperatures were tested to optimize recovery. The terpinen-4-ol recovery was unaffected by temperature and collection was carried out at an HE temperature of 35 °C. The linalool recovery, however, was optimized at a lower HE temperature of 25 °C. Percent recoveries at the optimized conditions (average yields are in the table) and chromatograms of the fractions can be viewed in Figure 3. Three sets of stacked collections (ten 100- $\mu$ L injections) were done for each compound under the optimized conditions. Based on the volatile nature of these compounds and the fairly low recoveries typically reported, 70–80% recovery was notable. Also, all fractions in the recovery study exhibited >99% enantiomeric purity.

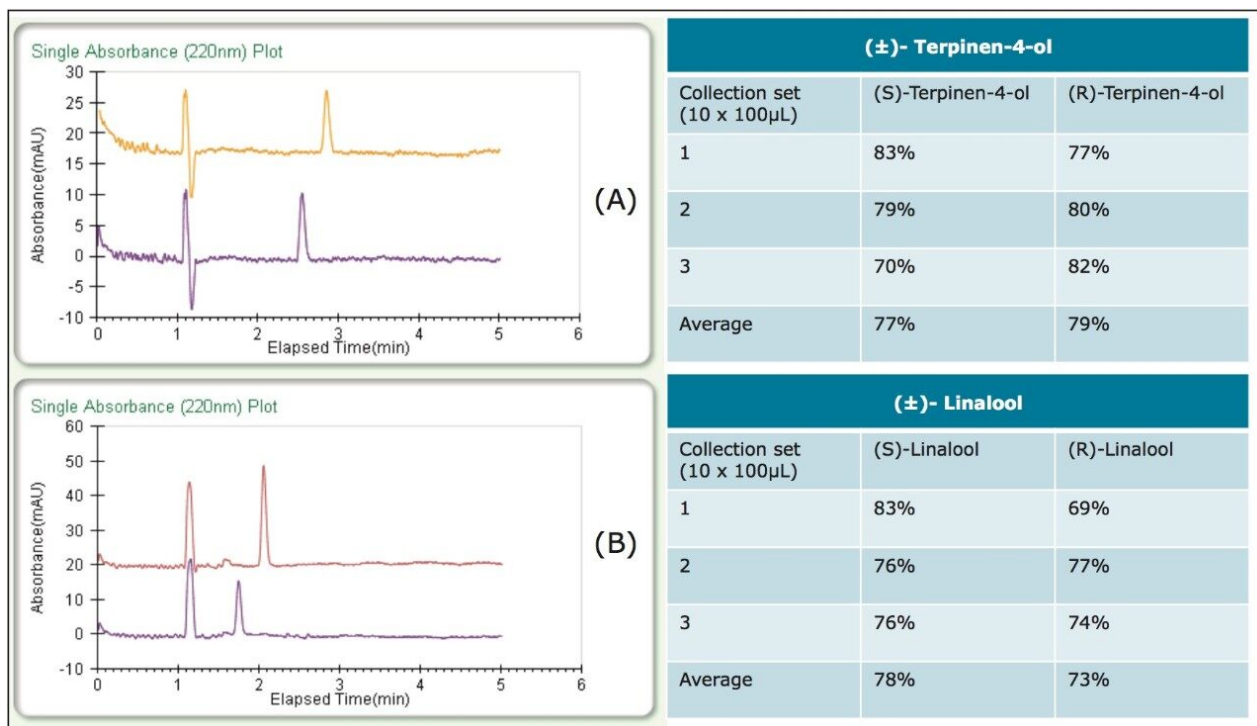


Figure 3. Chromatograms of the purified enantiomers of (A) ( $\pm$ )-terpinen-4-ol and (B) ( $\pm$ )-linalool

## Essential oil purifications

To test the purification technique with real samples, (R)- and (S)-terpinen-4-ol and (R)-linalool were purified from tea tree and lavender essential oils, respectively. Some method development was necessary to separate the enantiomers from their matrices, and to improve loading. The resulting methods were isocratic, allowing for stacked injections, and increasing collection efficiency. Due to the lower %content in the essential oils as compared to the standards, much higher loading was needed to get an acceptable yield for the compounds of interest. As a result, the samples were made up at much higher concentrations, 50 mg/mL for the tea tree oil, and 30 mg/mL for the lavender oil.

Figure 4 shows the separations and collection for the two essential oil purifications. The collected fractions were re injected directly without dilution; the resulting purity analysis can be seen in figure 5. The three target peaks were isolated resulting in >99% enantiomeric purity, and the overall chemical purity was also quite good (>92%).



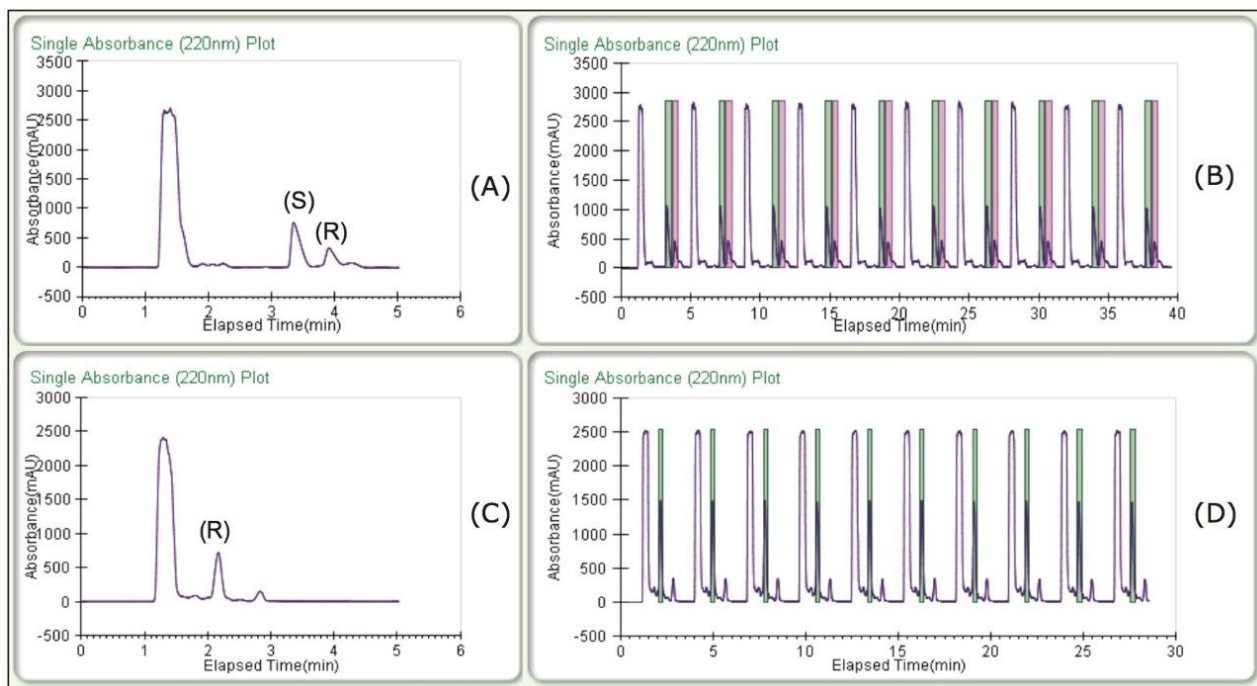


Figure 4. Preparative separations and stacked collections of terpinen-4-ol and linalool from tea tree oil (A and B) and lavender oil (C and D).

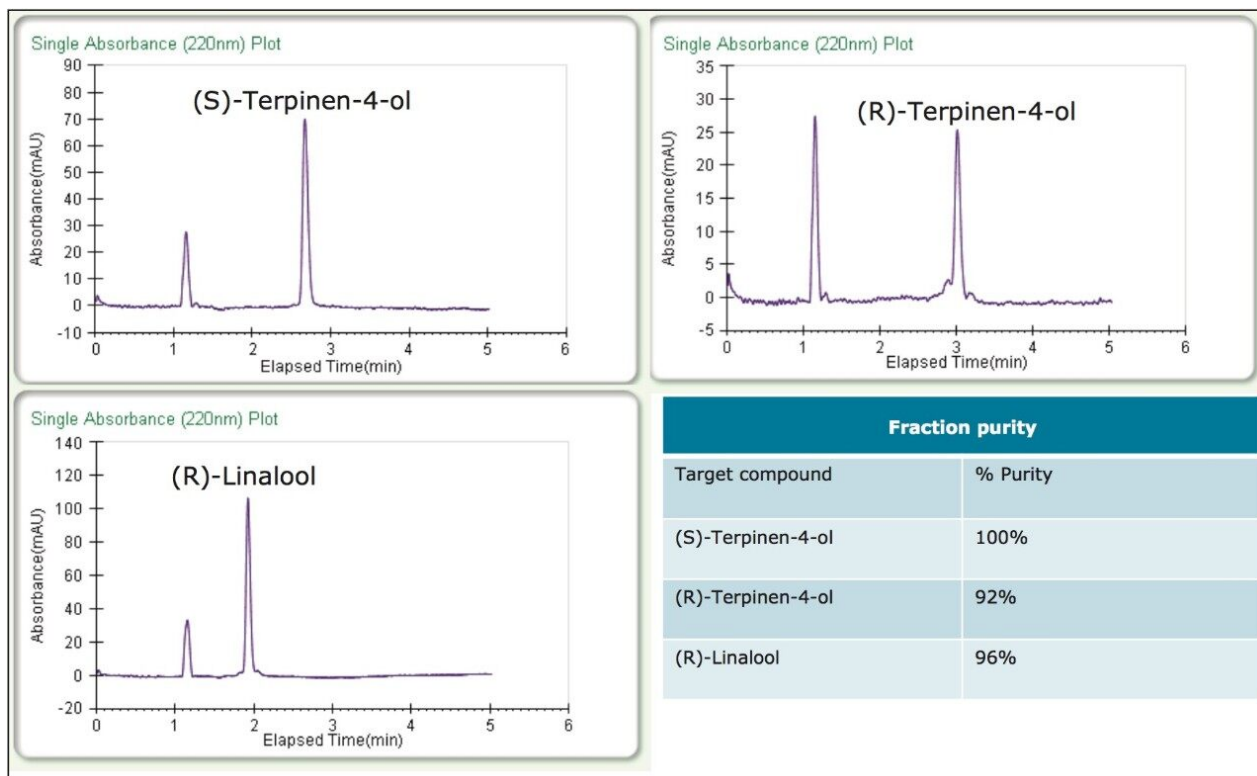


Figure 5. Chemical purity analysis of the tea tree and lavender essential oil fractions.

## Conclusion

- The Investigator SFC System employed a simplified chiral purification scheme for flavor and fragrance compounds without the need for complex sample trapping necessary in preparative GC.
- Approximately 10 mg of sample was purified in 30 minutes, resulting in enantiomerically pure fractions. This improved upon the efficiency of preparative GC, and the specificity of other purification techniques such as SPE or distillation.
- The low temperatures used in the Investigator SFC System conditions were shown to be suitable for flavor and fragrance compounds, leading to improved recoveries.
- Fast isocratic separations allowed for the use of stacked injections on the Investigator SFC System, resulting in higher throughput.
- Single-step chiral purification was achieved, resulting in good purity for the target compounds.

- A non-toxic, CO<sub>2</sub>:ethanol mobile phase was used, which is required for compounds destined for human use or consumption.

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## References

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2998 Photodiode Array (PDA) Detector <<https://www.waters.com/1001362>>

ChromScope Software <<https://www.waters.com/134647658>>

720005150, August 2014

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