

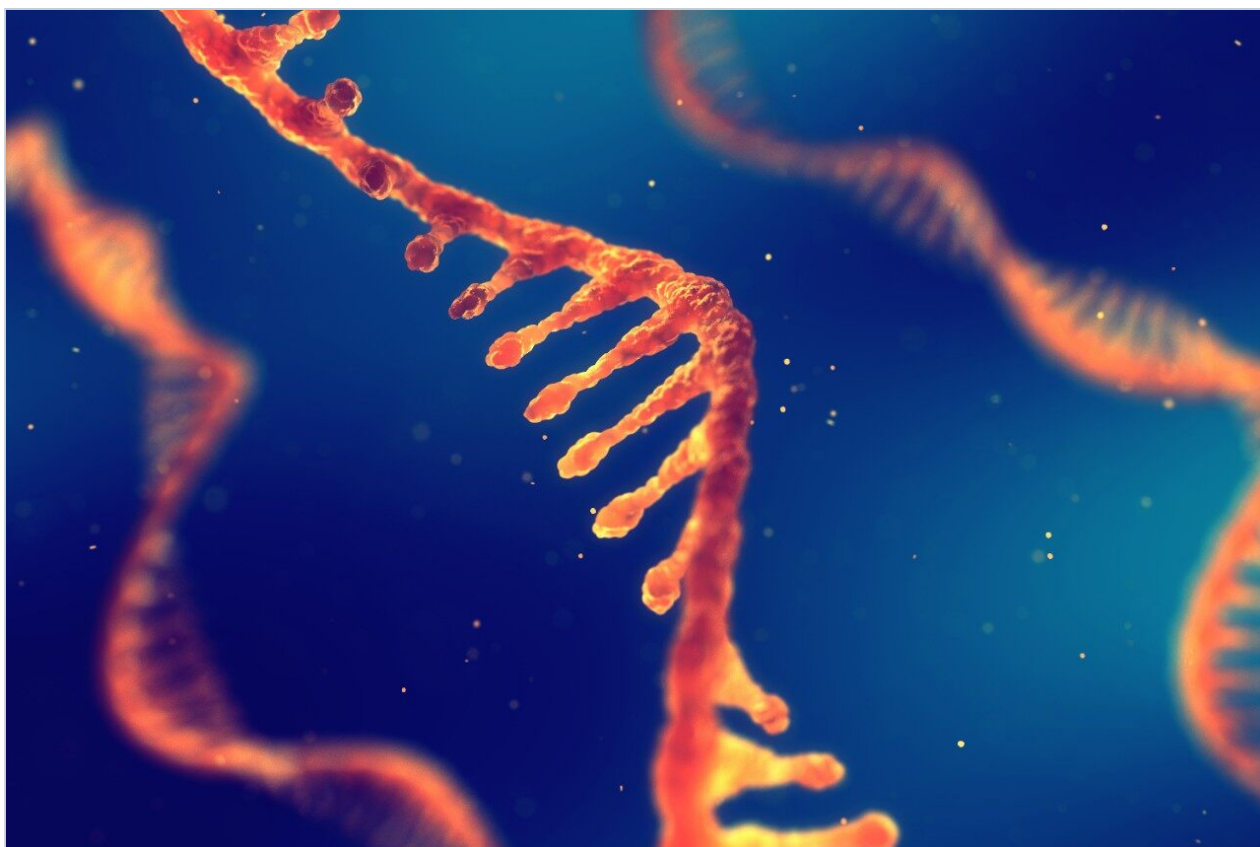
Application Note

## Hexylammonium Acetate as an Ion-Pairing Agent for IP-RP LC Analysis of Oligonucleotides

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Sean M. McCarthy, Martin Gilar

Waters Corporation



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## Abstract

In this application note, we outlined the use of hexylammonium acetate (HAA) for a variety of oligonucleotide chromatographic applications.

### Benefits

In this application note we describe the preparation of hexylammonium acetate (HAA) and the benefits to its use for a variety of applications compared to TEAA and/or TEA-HFIP for the chromatographic analysis of oligos, including:

- Homomolecular oligonucleotides
- Long oligonucleotides
- Heteromolecular oligonucleotides
- Phosphorothioates
- Labeled oligonucleotides
- Oligonucleotide duplexes

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## Introduction

Analysis of oligonucleotides via ion-pairing reverse-phase liquid chromatography requires efficient ion-pairing systems that yield chromatographic separation primarily based on charge/length.<sup>1</sup> The most commonly used ion-pairing agents are triethylammonium acetate (TEAA) and triethylamine hexafluoroisopropanol (TEA-HFIP). TEAA is commonly used due to its low cost and duplex compatibility, but suffers from limited resolution of oligonucleotides. TEA-HFIP offers benefits over TEAA, such as increased resolution and predictable retention.

The benefits are the result of increased ion-pairing and a decrease in reversed-phase behavior resulting from the nucleobases, evident in single-stranded oligonucleotide separations. Despite these benefits, the utility of a TEA-HFIP system is somewhat limited. In particular limitations arise for separation of duplex oligonucleotides, because it is denaturing and contributes significantly to melting. The costs associated with the use of TEA-HFIP are significantly higher than those associated with using acetate-buffered mobile

phases when considering purchase and disposal.

To overcome some of these issues, we investigated an ion-pairing system which uses hexylammonium acetate (HAA). HAA, when used with the Waters ACQUITY UPLC System, provides the same or better resolution of a variety of oligonucleotides, and is non-denaturing for duplexes. The cost associated with the use of HAA is significantly lower than that of HFIP-based mobile phases.

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## Experimental

### Mobile phase preparation

Hexylamine and acetic acid were purchased from Sigma Aldrich. The concentration of HAA in this example is 100 mM single-stranded; however, we have prepared 50 and 10 mM for increased MS-compatibility with little loss in resolution.

1 L of 100 mM HAA is prepared as follows:

It is highly recommended that mobile phase components be measured by mass instead of volume. Significant variation in retention times may result from inaccurate mobile phase preparation.

1. Weigh 981 g of water and place in a beaker with a Teflon coated magnetic stir bar.
2. Add 5.99 g (5.71 mL) of acetic and stir the mixture.
3. Add 10.12 g (13.21 mL) of hexylamine.
4. Stir the resulting solution so there is a significant vortex for 5 minutes.
5. The pH of the solution should be approximately 7.0. If the pH needs to be adjusted, carefully add acetic acid or hexylamine as appropriate in  $\sim 1$   $\mu$ L aliquots and stir until dissolved. This solution is used as MP A.
6. MP B is prepared as 50% ACN in MP A. This should be prepared by mass to ensure reproducible results. The density of MP A is assumed to be the same as water for the purpose of MP B preparation.

The mobile phase can be used for several days without any significant loss in oligonucleotide resolution.

Oligonucleotide samples should be dissolved in either 100 mM HAA or 100 mM TEAA, pH 7.0. There is strong affinity of hexylamine for the particle sorbents; therefore, it is highly recommended that a column be

dedicated for use with HAA due to its high affinity for the stationary phase.

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## Results and Discussion

### Homomolecular oligonucleotides

The Waters Oligonucleotide Separation Technology (OST) standard demonstrates the resolving capabilities of HAA compared to traditionally-used mobile phases. The separations shown in Figure 1 were normalized by adjusting each gradient so that the 15-mer and 35-mer peaks eluted at the same time with each mobile phase system. As shown, HAA offers a significant increase in resolution compared to the commonly-used, non-denaturing TEAA mobile phase. The increase in resolution is attributed to the increased ion pairing ability of HAA. Comparing the separation with HAA to the separation with TEA-HFIP clearly indicates that HAA provides similar or better resolution than HFIP-based mobile phase. In fact, our data indicated that the resolution of longer oligonucleotides is better with HAA.

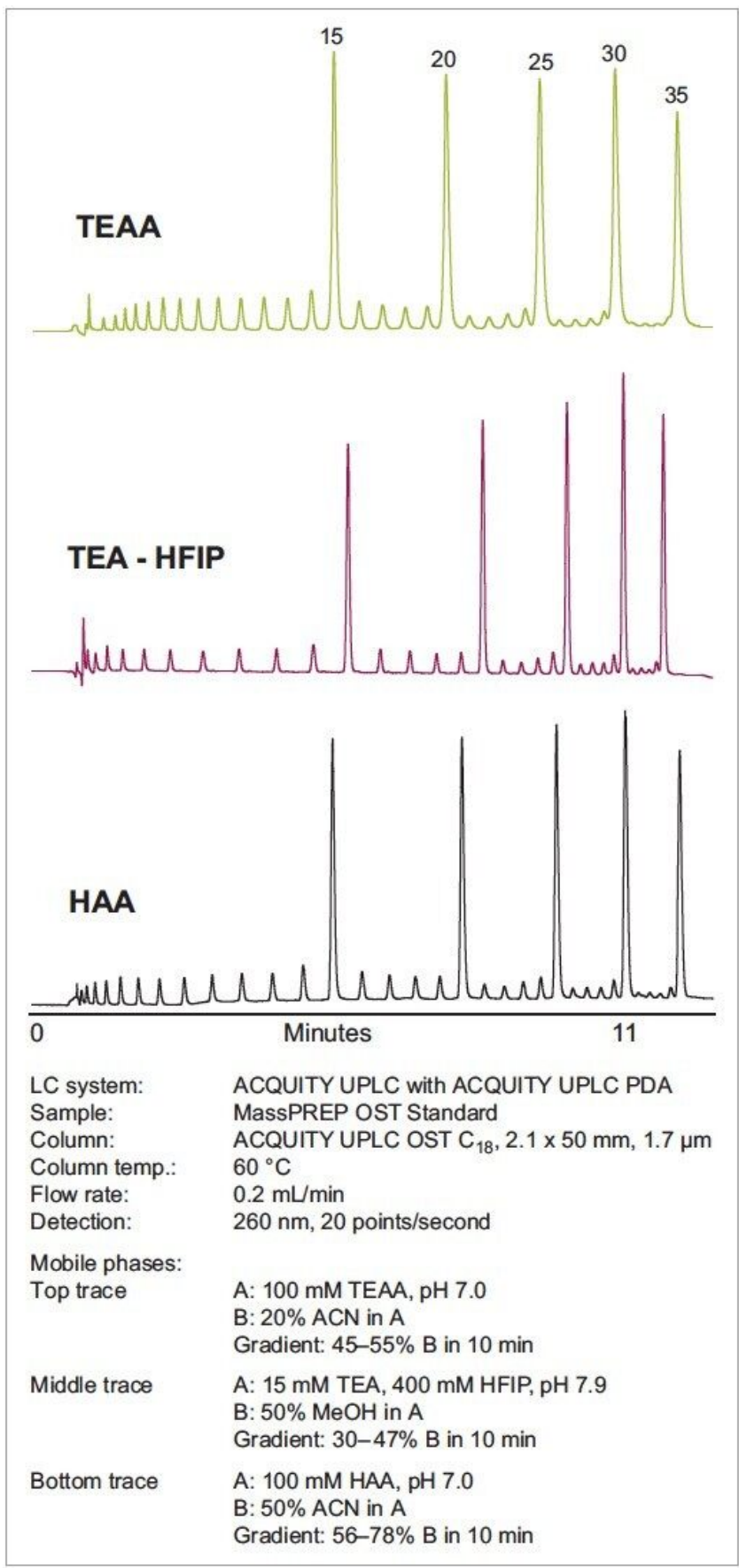


Figure 1. Separation of MassPREP OST standard with different ion-pair

Long oligonucleotides

be very challenging. This is largely due to the inherent chirality of the phosphorothioate linkage, which often leads to broad peaks and incomplete resolution of truncated sequences due to isomeric distribution. Figures 4 and 5 show that HAA adequately resolves truncated phosphorothioate sequences from the full-length sequence. The ability to resolve N-1 phosphorothioates indicates that HAA partially suppresses diastereomeric resolution. While the MS sensitivity resulting from the use of HAA is lower than that of TEA-HFIP, useful MS data is easily obtained and confirms the separation of the full-length phosphorothioate from truncated sequences as shown in the selected ion chromatograms in Figure 5.

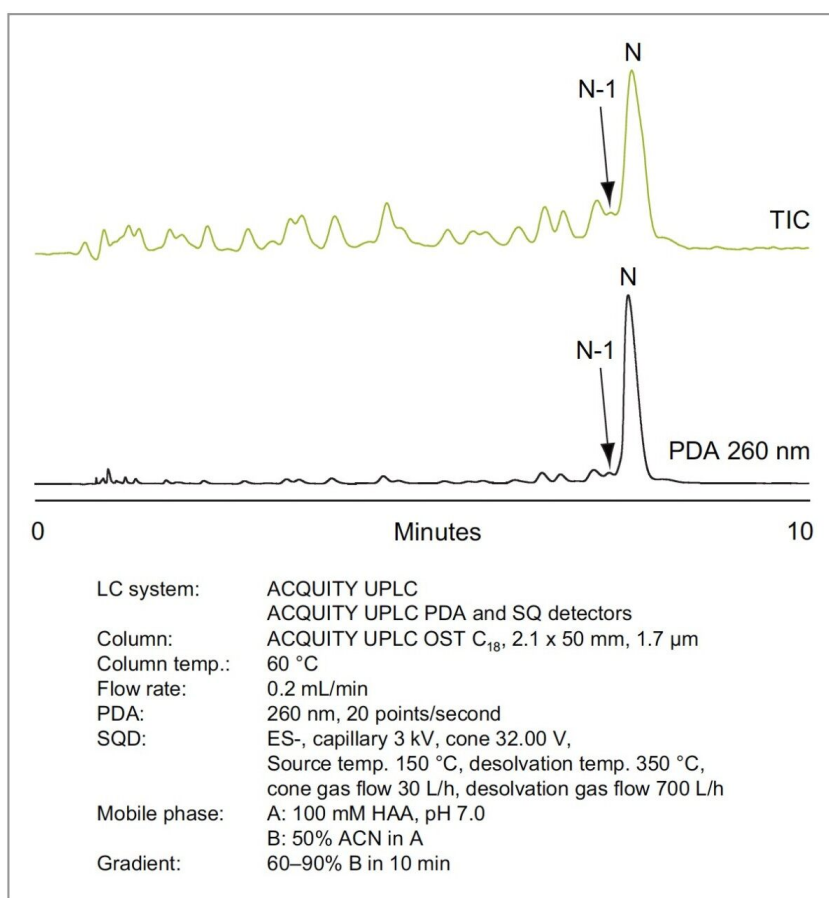


Figure 4. Separation of phosphorothioate with HAA.

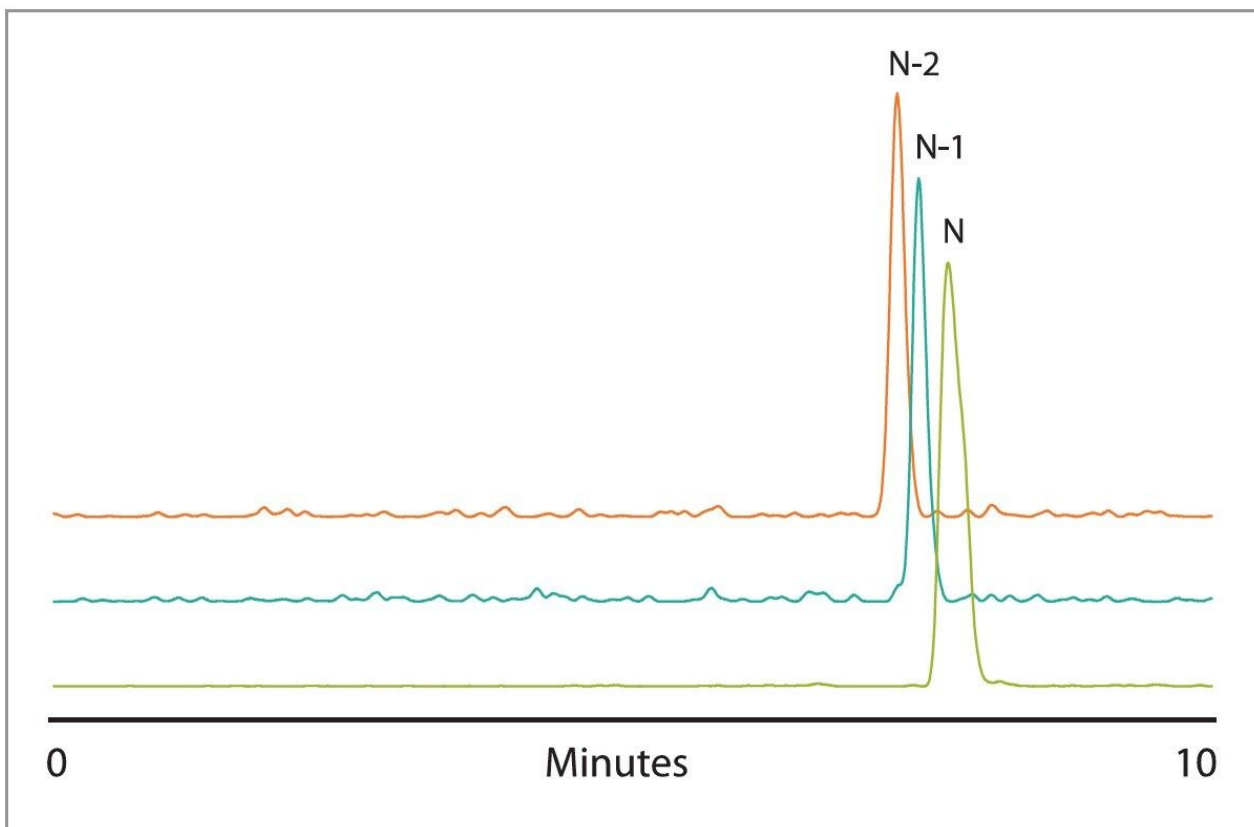


Figure 5. Selected ion chromatograms of phosphorothioate.

### Labeled oligonucleotides

Many oligonucleotides are labeled with fluorescent tags for a variety of reasons. These tags are generally large aromatic and poly-aromatic groups that impart significant hydrophobicity to the oligonucleotide. For this reason, inefficient ion pairing often leads to poor resolution of labeled species as hydrophobic forces overcome the ion-pair mechanism. Figure 6 illustrates the analysis of a hydrophobically-labeled oligonucleotide with TEA-HFIP (top trace) and HAA (bottom trace). TEA-HFIP does not yield the desired elution pattern. Instead, the target peak is followed by truncated labeled species. The reversal in elution order results from increased contribution of the hydrophobic tag to chromatographic behavior as the hydrophilic oligonucleotide size decreases. The use of a more efficient ion-pairing agent, such as HAA, results in regular retention of all labeled species in their expected order.

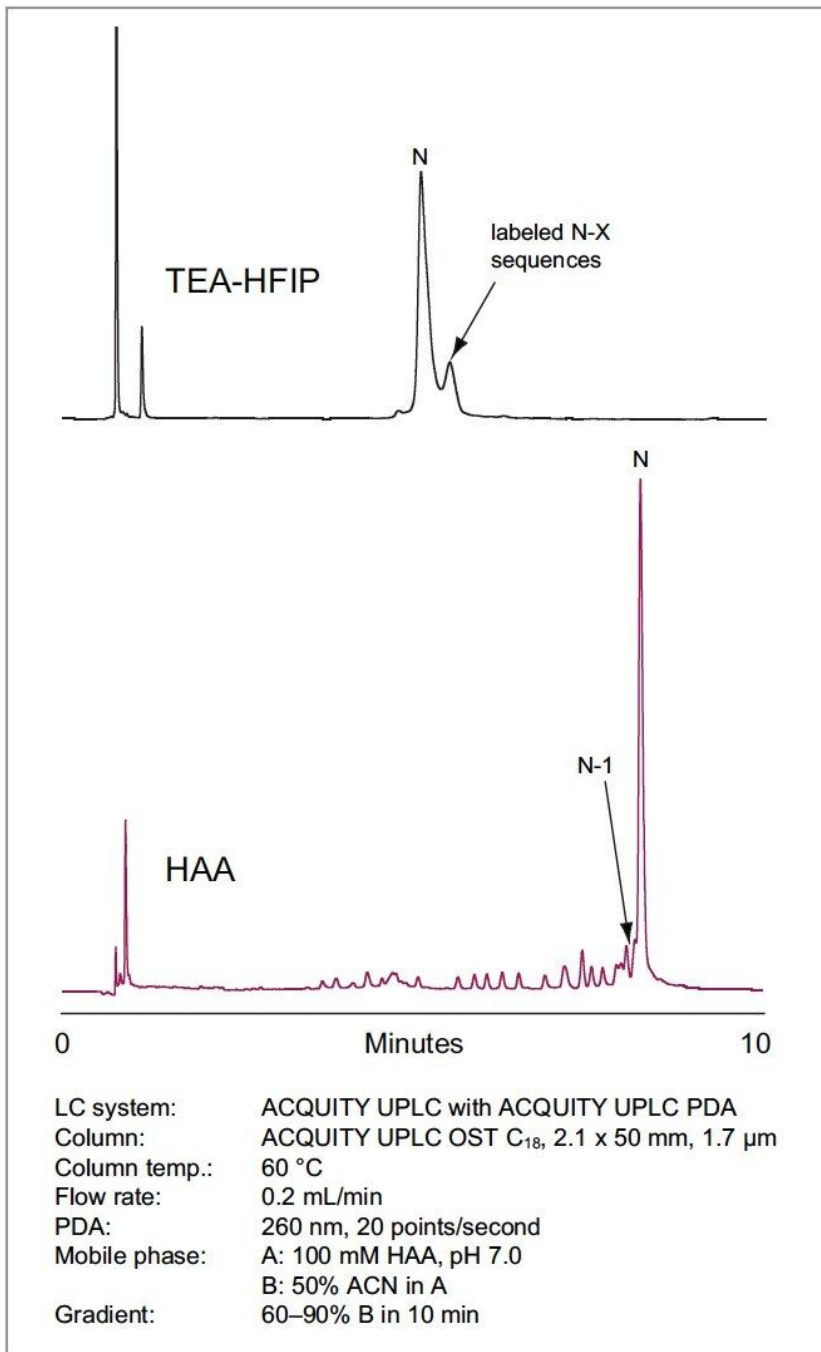


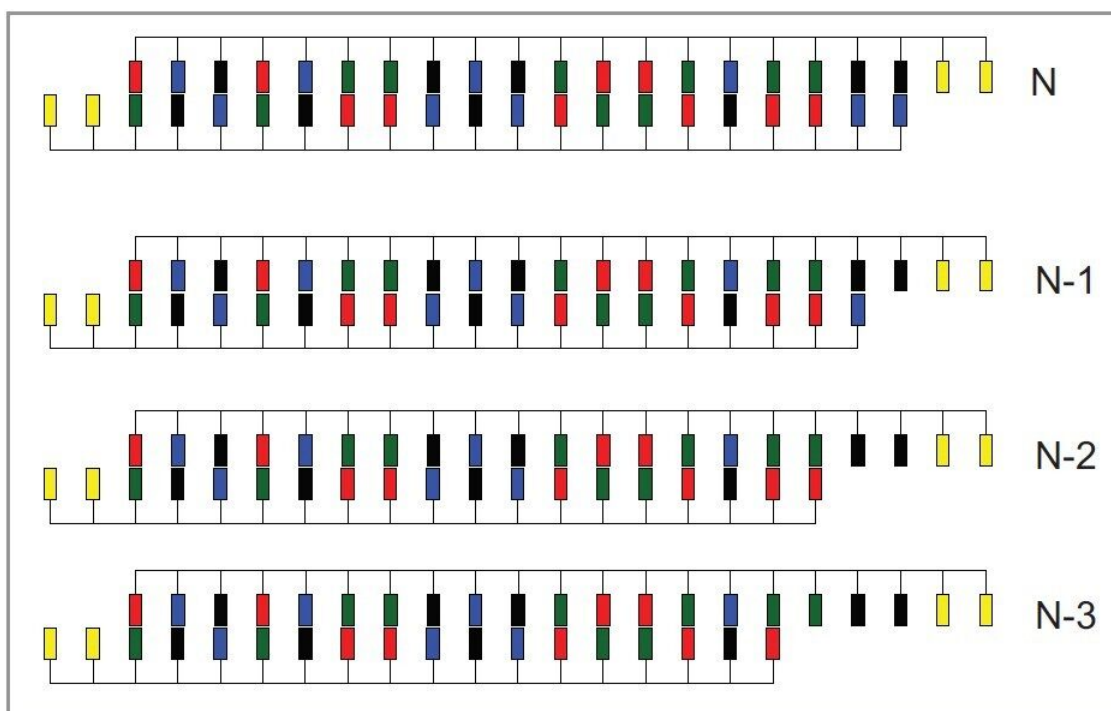
Figure 6. Separation of labeled oligonucleotide.

## Oligonucleotide duplexes

As mentioned previously, HAA is non-denaturing and is fully compatible with oligonucleotide duplexes. We investigated the ability of HAA to resolve mismatched from full-length RNA duplexes. To demonstrate the ability of HAA to resolve RNA duplexes, we utilized a purified upper RNAi strand and a partially-digested



lower strand. Upon annealing, a ladder of siRNA duplexes is formed with partially 5' truncated lower RNAi strands (Figure 7).



*Figure 7. Duplexes formed by annealing full-length upper RNAi strand with partially-truncated lower RNA strand.*

As shown in Figure 8A, HAA successfully resolves truncated siRNA duplexes from full-length duplex and single-stranded RNAi species. Using hexylammonium acetate we were also able to identify each eluting duplex peak by the corresponding mass of complementary RNAi strands with MS. In this way we were able to confirm the elution order of the impurities. Extracted selected ion chromatograms shown in Figure 8B indicate resolution of truncated duplexes.

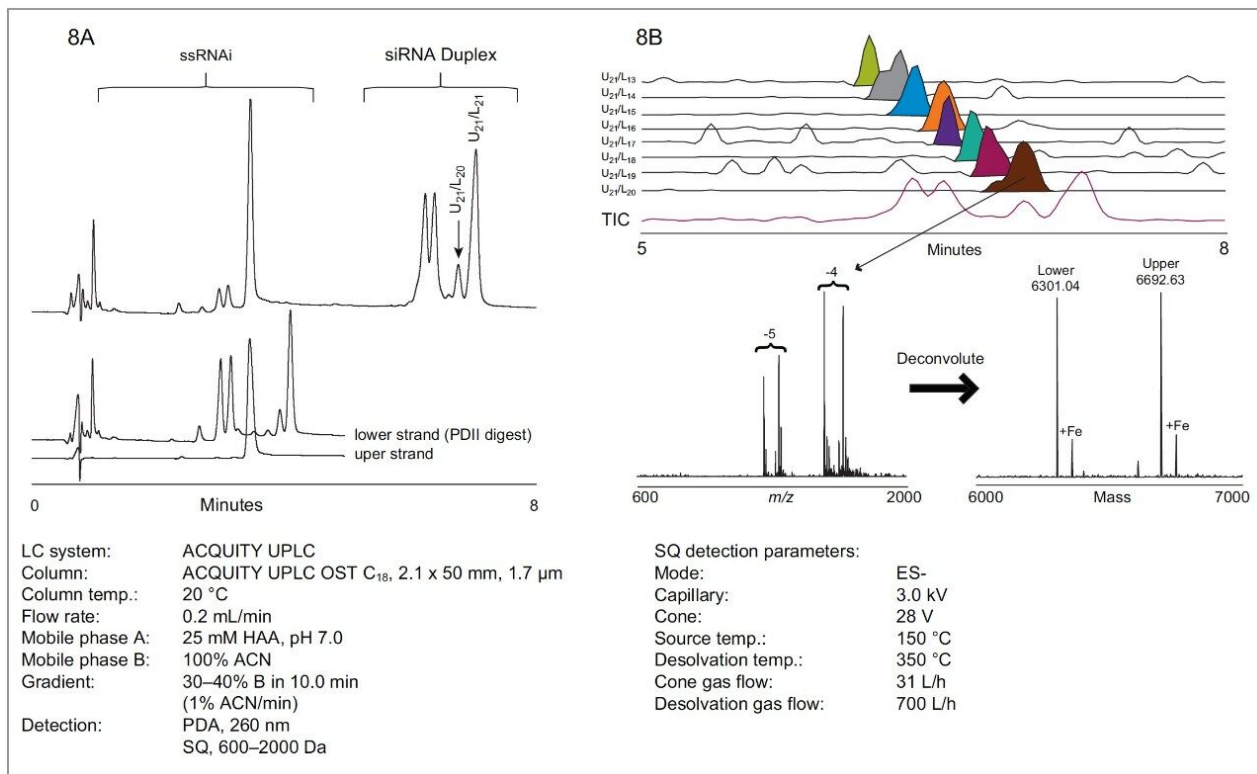


Figure 8. Separation of siRNA duplexes and single stranded RNA.

## Conclusion

In this application note, we outlined the use of HAA for a variety of oligonucleotide chromatographic applications. HAA and TEA-HFIP exhibit similar resolution of moderate length, unmodified homo and heteromolecular oligonucleotides. For separation of longer oligonucleotides, ca. >35-mer, HAA exhibits better resolution which can be further improved with longer columns and correspondingly longer gradient times. HAA is able to adequately resolve phosphorothioates, a particularly difficult separation. The separation of labeled oligonucleotides shows a clear advantage of HAA over TEA-HFIP with HAA giving predictable retention of labeled species and minimal contribution of the hydrophobic label. Finally, the non-denaturing character of HAA allows for its use with oligonucleotide duplexes and offers impressive resolution of mismatches. While HAA does not offer the same MS compatibility as TEA-HFIP, its utility is evident. Taken together this data indicates that the use of HAA with the ACQUITY UPLC System provides exceptional separation of a variety of oligonucleotides at a significantly lower cost than TEA-HFIP.

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## References

1. Oligonucleotide Separation Technology XBridge OST C<sub>18</sub> Columns Method Guidelines. Waters User Manual. 2008. 715001476.

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