

## SPE-LC/MS Bioanalytical Quantification of the Biotherapeutic Peptide, Semaglutide From Plasma

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### Abstract

The following work demonstrates the sensitive, selective, and robust LC-MS bioanalytical quantification of the peptide therapeutic, semaglutide from plasma. Using a combination of selective solid-phase extraction (SPE) sample preparation in the  $\mu$ Elution 96-well format, QuanRecovery™ with MaxPeak™ High Performance Surfaces, 96-well plate for sample analysis to mitigate peptide non-specific binding, and LC-MS/MS analysis and quantification using the UPLC™ Peptide CSH™ C<sub>18</sub> Column, ACQUITY™ UPLC I-Class Plus System, and Xevo™ TQ-XS Mass Spectrometer, to achieve fast, sensitive, and highly reproducible and accurate quantification of semaglutide from plasma biomatrices.

### Benefits

- Selective, fast SPE extraction (<30 minutes)
  - 96-well  $\mu$ Elution plate format for SPE enables concentration of the sample while maintaining solubility and minimizes peptide loss due to adsorption
  - Fast UPLC analysis (4-minutes) using the ACQUITY™ Peptide CSH C<sub>18</sub> Column
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- Use of QuanRecovery with MaxPeak HPS sample collections plates, mitigated non-specific binding of Semaglutide, ensuring high peptide recovery, and repeatability during analysis
- Accurate and sensitive quantification of semaglutide with an LLOQ of 0.120 ng/mL from extracted plasma

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## Introduction

Semaglutide (Ozempic®, Rybelsus®, Wegovy®) is a 32 amino acid, glucagon-like-peptide 1 (GLP-1) analog used in the treatment of type 2 diabetes and as a treatment for obesity.<sup>1</sup> With continued research and development of this peptide and closely related GLP-1 agonist peptides, there is continued need for their robust, sensitive, and selective sample preparation and liquid chromatography mass spectrometry (LC-MS) analysis. However, sensitive and robust quantification of large peptides, like semaglutide (MWT 4114), are often difficult to analyze by LC-MS/MS, as MS sensitivity is low due to poor transfer of the peptide into the gas phase, dilution of signal across charge states, and poor fragmentation. In addition, the semaglutide suffers from significant non-specific binding and poor solubility, making LC and sample preparation method development challenging.

The work described herein takes advantage of a selective  $\mu$ Elution reversed-phase solid-phase-extraction (SPE), optimal MS settings and MRM transitions, and a high-efficiency UPLC separation using a column packed with C<sub>18</sub> particles containing a low-level positive surface charge to provide excellent chromatography peak shape, and use of a low binding sample collection plate to mitigate hydrophobic peptide loss, ultimately facilitating highly sensitive, and robust, low-ng/mL quantification of semaglutide from plasma.

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## Experimental

### Sample Preparation

Semaglutide (Cayman Chemical) was added to commercially available rat plasma. Calibration curve samples concentrations were between 0.120–1000 ng/mL. Semaglutide quality control (QC) samples were prepared at 0.48, 15.6, and 500 ng/mL. Standard curve and QC plasma samples were prepared in triplicate in protein low bind 1.5 mL polypropylene tubes.

Step 1: Protein precipitation (PPT)

200  $\mu\text{L}$  aliquots of the prepared semaglutide rat plasma samples were precipitated with 200  $\mu\text{L}$  methanol and centrifuged for 5 minutes @ 10,000 rcf.

The supernatant was transferred to a 1 mL QuanRecovery 96-well plate containing 400  $\mu\text{L}$  of water and mixed.

Step 2: SPE using an Oasis HLB 96-well  $\mu\text{Elution Plate}$

A 3-step (load, wash, and elute) SPE protocol was employed. The entirety (600  $\mu\text{L}$ ) of the diluted PPT supernatant samples were loaded onto the extraction plate, followed by a 200  $\mu\text{L}$  wash with 5% methanol. A 2x25  $\mu\text{L}$  elution of the purified sample was performed using a 75/25 acetonitrile/water solution containing 5% formic acid, collected in a QuanRecovery 96-well collection plate. Subsequent analysis of the SPE extracted sample was performed.

## LC/MS Conditions

LC system:	ACQUITY UPLC I-Class Plus, Fixed-Loop
Mobile phase A:	0.1% Formic acid in water
Mobile phase B:	0.1% Formic acid in acetonitrile
Weak needle wash:	90:10 Water:Acetonitrile with 0.1% formic acid
Strong needle wash:	Acetonitrile:Isopropanol:Water:Methanol (25:25:25:25 v/v/v/v)
Detection:	Xevo TQ-XS
Column(s):	ACQUITY UPLC Peptide CSH C <sub>18</sub> 100 Å, 1.7 $\mu\text{m}$ 2.1 mm x 50 mm (p/n: 186005296)
Column temperature:	65 °C
Sample temperature:	10 °C

Injection volume: 10  $\mu$ L

Flow rate: 0.3 mL/min

## LC Gradient

Time (min)	Flow (mL/min)	%A	%B	Curve
Initial	0.300	65.0	35.0	6
0.25	0.300	65.0	35.0	6
3.0	0.300	25.0	75.0	6
3.1	0.300	10.0	90.0	6
3.2	0.300	10.0	90.0	6
3.4	0.300	65.0	35.0	6

## MS Settings

Ionization mode: ESI+

Acquisition range: MRM

Capillary voltage: 2.00 kV

Cone voltage: 32 V

Desolvation temperature: 500  $^{\circ}$ C

Desolvation flow: 1100 L/Hr

Cone gas flow: 150 L/Hr

Collision gas flow: 0.15 mL/min

Nebulizer gas flow: 7 Bar

## Data Management

Instrument control software: MassLynx™ (v4.2)

Quantification software: TargetLynx™

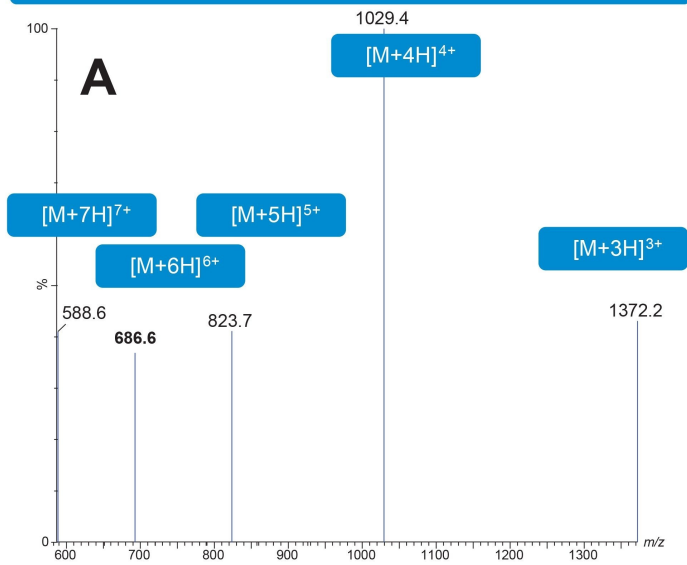
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## Results and Discussion

LC-MS/MS quantification of semaglutide was performed using an ACQUITY UPLC I-Class coupled to the Xevo TQ-XS MS System operated in ESI+ mode. During MS method development, multiply charged precursors between  $[M+3H]^{3+}$  to the  $[M+7H]^{7+}$  of semaglutide were observed performing a direct MS infusion of a semaglutide solution (Figure 1A). Following infusion, confirmation of these charge states under LC chromatographic separation and MS detection using Selected Ion Monitoring (SIM) was performed. Under these conditions it was determined that the most sensitive precursors were  $[M+4H]^{4+}$  with an  $m/z$  of 1371.2 and the  $[M+3H]^{3+}$  precursor with an  $m/z$  of 1029 (Figure 1B). During optimization for MRM fragmentation identification, it was found that the  $[M+3H]^{3+}$  precursor was harder to fragment, requiring a very high collision energy, and while it had a high MS sensitivity in SIM mode, the fragments evaluated for MRM analysis were > two orders of magnitude less sensitive than the  $[M+4H]^{4+}$  precursor and its resulting fragments (Figure 2). For this reason, final MS/MS multiple reaction monitoring (MRM) transitions used for detection and quantification, were 1029.27>1302.6 use for primary quantification, while the 1029.27>1238.1 and 1371.2>1238.1 MRMs were used for confirmation.

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Semaglutide MS spectra and representative SRM chromatogram



## Semaglutide MS Spectra and Representative SIM Chromatogram

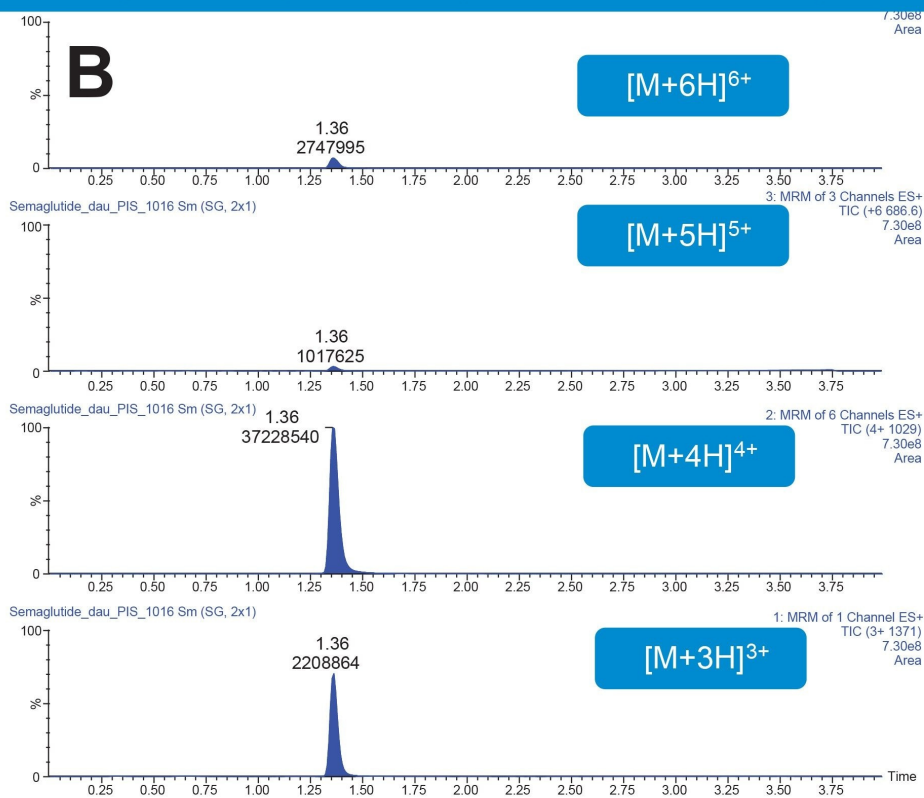


Figure 1. MS spectra (A) highlighting the  $[M+3H]^{3+}$  -  $[M+7H]^{7+}$  multiply charged precursors of semaglutide and representative single reaction monitoring (SRM) chromatograms highlighting MS response for the  $[M+3H]^{3+}$  -  $[M+6H]^{6+}$  multiply charged precursors of semaglutide (B).

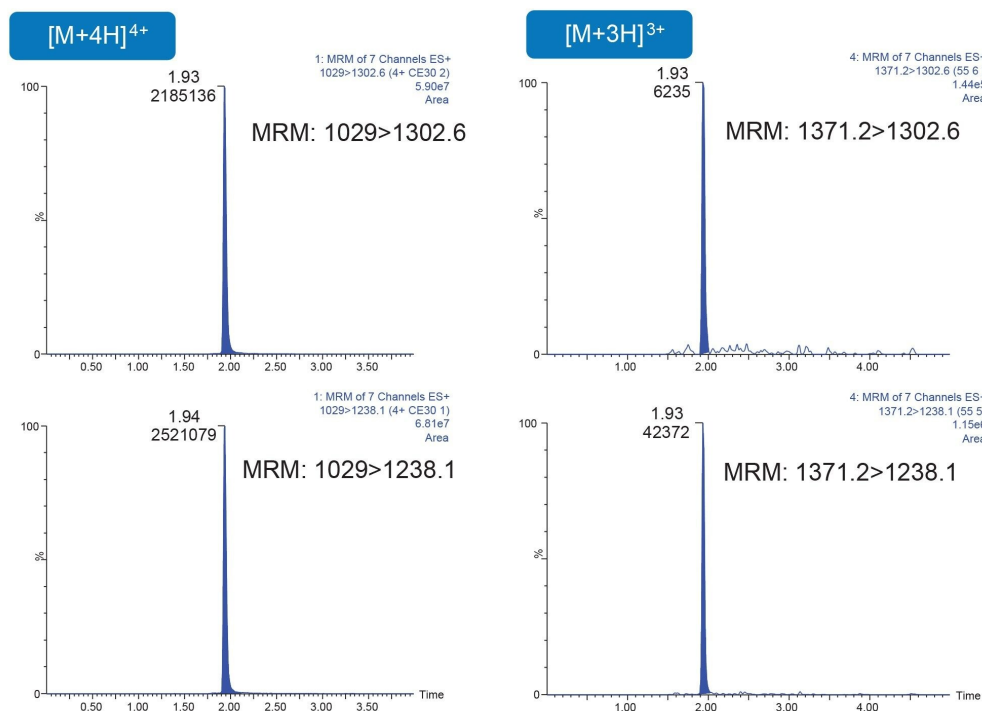


Figure 2. Representative semaglutide MS MRM chromatograms of the  $[M+3H]^{3+}$  and  $[M+4H]^{4+}$  multiply charged precursors, highlighting best sensitivity using the  $[M+4H]^{4+}$  multiply-charged precursor and the 1302.6 and 1238.1  $m/z$  fragments.

Semaglutide is a large and hydrophobic peptide with a molecular weight of 4113.5 g/mol and HPLC Index of 89.7 (relative measure of hydrophobicity). Chromatography of large hydrophobic peptides is often challenged with poor peak shape, peak tailing, and peptide carry-over, with root causes often associated with poor solubility, poor diffusion in and out of the chromatographic pore, and sub-optimal column temperature. To alleviate these issues, column temperatures were maintained at 65 °C, and lower flow rates (between 0.3–0.5 mLs/min) were employed. Using lower flow rates and elevated column temperature improved semaglutide retention time, facilitating diffusion of semaglutide in and out of the chromatographic pores, while elevated temperatures reduced peak tailing and minimized column carry-over. One of the biggest improvements with chromatography was in the evaluation of column sorbents. This evaluation focused on an ACQUITY UPLC Peptide BEH C<sub>18</sub> (300 Å) 1.7 µm Column and the CSH ACQUITY UPLC Peptide CSH C<sub>18</sub> Column (130 Å) 1.7 µm Column, both well-documented for excellent peptide separation performance. The CSH column packed with C<sub>18</sub> particles containing a low-level positive surface charge provided superior chromatography, with a better MS response and reduced



peak tailing of semaglutide. This performance is illustrated in Figure 3. For these reasons, the CSH C<sub>18</sub> column was used for semaglutide quantitative analysis.

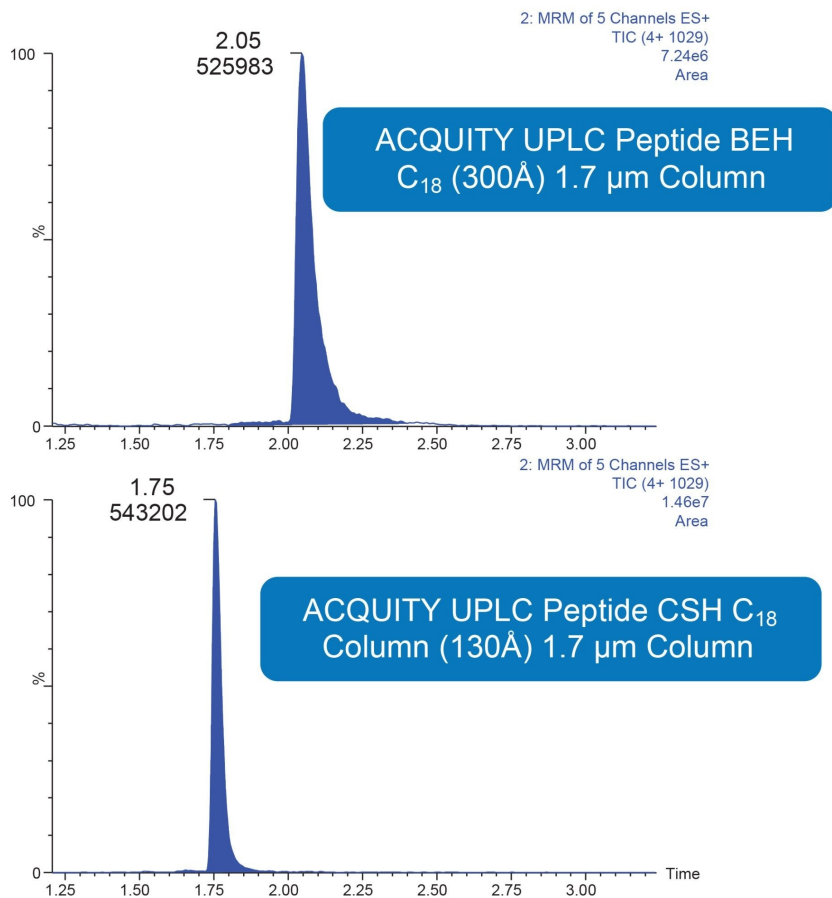
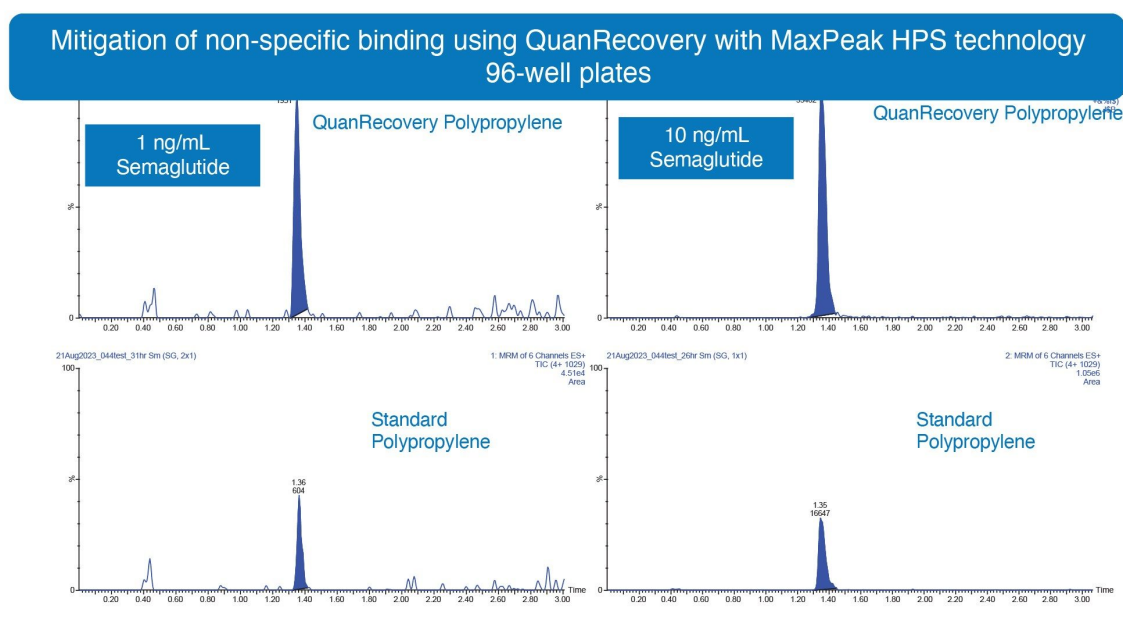


Figure 3. Semaglutide LC column evaluation, comparing the ACQUITY UPLC Peptide BEH C<sub>18</sub> (300 Å) 1.7 µm Column and the ACQUITY UPLC Peptide CSH C<sub>18</sub> Column (130 Å) 1.7 µm Column (0.4 mL/min @ 65°C), and demonstrating increased MS response and reduced peak tailing with the Peptide CSH C<sub>18</sub> Column.

An additional challenge working with semaglutide was encountered during sample preparation and SPE purification due to a high degree of semaglutide non-specific binding (NSB) to consumables and difficulty maintaining peptide solubility throughout this process. As part of the sample preparation and extraction protocol

development, consumables used during sample preparation and LC-MS analysis were evaluated, with key focus on polypropylene consumable designed specifically to mitigate NSB due to peptide hydrophobic interaction. In this evaluation, standard polypropylene and QuanRecovery 96-well sample plates were compared. QuanRecovery plates, with MaxPeak high performance surfaces (HPS) technology were designed specifically to minimize peptide-surface interaction mitigating hydrophobic NSB loss. This comparison, highlighting improved MS response using QuanRecovery vs standard polypropylene plates, using semaglutide solutions (one and ten ng/mL) prepared in a 10% methanol solution, is highlighted in Figure 4. For the same reasons, low-bind polypropylene tubes were also used to further mitigate NSB loss during semaglutide stock solution preparation. Additionally, it was found that best the best MS signal was seen for semaglutide stock solutions prepared in a 20–30% methanol solution vs 100% aqueous or 100% methanol solutions.



*Figure 4. Mitigation of hydrophobic NSB to collection vessels using QuanRecovery™ with MaxPeak HPS 96-well Sample Collection plate, demonstrating improved MS peak response vs standard polypropylene plates.*

Sample pretreatment prior to SPE purification proved to be critical in improving recovery and specificity of semaglutide extraction from plasma. Traditional sample pretreatments of acidic and/or basic aqueous solutions, failed to adequately disrupt semaglutide-plasma protein binding, yielding <20% recovery from plasma.

Employing a protein precipitation (1:1) with 5% formic acid methanol pretreatment resulted in 80–100% recovery semaglutide without precipitating the peptide itself. Protein precipitation with higher ratios of organic resulted in peptide loss due to undesired precipitation of semaglutide. Additionally, this, PPT pretreatment further eliminated endogenous interferences from large proteins such as albumin, adding selectivity to this SPE extraction. Following PPT sample pretreatment, the semaglutide PPT supernatant was diluted 1:2 with water prior to loading onto the Oasis HLB SPE plates. This was done to minimize breakthrough of semaglutide due to high organic composition of the PPT sample. This ensured semaglutide was well retained on the SPE sorbent during the load step, with no breakthrough occurring. The optimum elution solution was determined to be 75% acetonitrile containing 5% formic acid, ensuring optimum elution while maintaining adequate solubility of semaglutide. This final protocol is described in the experimental section and illustrated in Figure 5. Quantitative performance for the calibration curve of semaglutide extracted from plasma is highlighted in Table 1, while QC performance is highlighted in Table 2 and illustrated in Figure 6.

#### **Oasis® HLB 96-well $\mu$ Elution Plate (2 mg/well)**

##### **Sample pretreatment**

1:1 200  $\mu$ L plasma: 200  $\mu$ L methanol PPT  
Dilute 1:2 with water prior to SPE loading

##### **Load**

Entirety diluted plasma PPT supernatant

##### **Wash**

1 x 200  $\mu$ L in 5% MeOH

##### **Elute**

2 x 25  $\mu$ L 75/25 acetonitrile/water with 5% formic acid  
Dilute with 50  $\mu$ L water (optional)

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*Figure 5. Semaglutide sample extraction protocol using a 1:1 plasma:methanol pretreatment followed by reversed-phase SPE using the Oasis HLB 96-well  $\mu$ Elution Plate (2 mg/well), providing 2x sample concentration using a 200  $\mu$ L starting plasma volume and a 50  $\mu$ L SPE elution.*

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Semaglutide rat plasma calibration curve statistics			
Calibration curve dynamic range (ng/mL)	Weighting	Linear fit (R <sup>2</sup> )	% Accuracy range
0.12–1,000	1/X <sup>2</sup>	0.995	-11.6–14.8

Tables 1. Calibration curve statistics of semaglutide extracted from plasma.

Semaglutide rat plasma QC statistics				
QC level	Expected concentration (ng/mL)	Mean observed concentration (ng/mL) (N=3)	Mean % accuracy (N=3)	% RSD
LQC	0.48	0.50	103.80	3.80
MQC	15.6	16.09	103.10	4.28
HQC	500	497.14	99.43	6.04

Tables 2. QC statistics of semaglutide extracted from plasma.

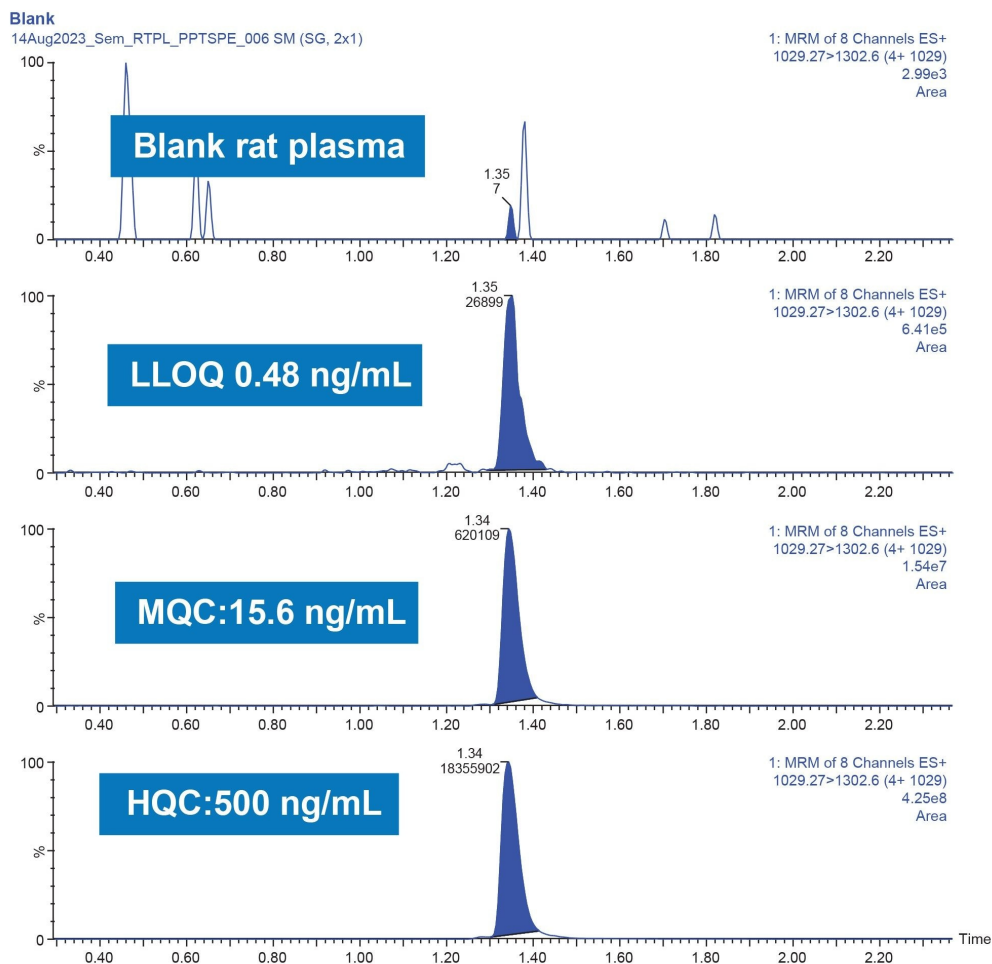


Figure 6. Representative chromatograms of semaglutide QC samples extracted from plasma at 0.48, 15.6, and 500 ng/mL.

## Conclusion

The combination of a selective reversed-phase  $\mu$ Elution SPE cleanup, mitigation of NSB with QuanRecovery MaxPeak HPS 96-well plates, proper MS fragment choice, and chromatographic separation using the Peptide CSH C<sub>18</sub> UPLC Column, enabled sensitive, accurate, and robust semaglutide quantification from plasma.

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## References

1. Semaglutide: Uses, Interactions, Mechanism of Action | DrugBank Online <<https://go.drugbank.com/drugs/DB13928>> , accessed on 19 September 2023.

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